Characterization of a novel vasorelaxing peptide derived from the G Protein-coupled Estrogen Receptor 1

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BACKGROUND

 Vascular endothelial cells are the physiological source of nitric oxide (NO), which is crucial for the control of vascular tone, cardiac functions, and other beneficial actions in many organ systems.

 Many endothelial functions are dependent on store-operated Ca²⁺ entry (SOCE) (Fig. 1A). Herein, depletion of the endoplasmic reticular (ER) Ca²⁺ content triggers intra-ER conformational changes in the stromal interaction molecule 1 (STIM1) to promote its interaction with plasma membrane channels for Ca²⁺ entry (Fig. 1B). Sustained ER Ca²⁺ depletion, however, is associated with ER stress and endothelial dysfunction, a common feature of many pathological conditions such as hypertension and diabetes.

 Our laboratory previously identified a specific interaction between the intra-ER Ca²⁺-binding loop of STIM1 and the G proteincoupled estrogen receptor (GPER), a receptor that mediates many cardiovascular protective functions (reviewed *Front Endocrinol* 2020, *PMID* 33133016). From these studies, we have developed a peptide (G2) based on the GPER-STIM1 interacting domain and conjugated it with an endothelium-specific leader sequence to generate a tandem peptide (EFG2). Due to the biochemical properties of the G2 domain, EFG2 is predicted to specifically promote endothelial Ca²⁺ entry and Ca²⁺-dependent functions without emptying ER Ca²⁺.



Figure 1. Role and mechanisms of SOCE in endothelial NO production and vasorelaxation. A, Benefits of endothelial SOCE and consequent NO production. B, Molecular mechanism of STIM1 involvement in activation of SOCE. PM, plasma membrane; ER, endoplasmic reticulum; CC1, coiled-coiled domain 1; CAD, CRAC [Ca²⁺ release-activating Ca²⁺ entry]-activating domain; Orai1, a key store-operated Ca²⁺ channel.

HYPOTHESIS

By specifically targeting endothelial cells and binding STIM1's Ca²⁺-binding loop with high affinity, EFG2 promotes STIM1's conformational changes that trigger Ca²⁺ entry in endothelial cells without causing ER Ca²⁺ release, thereby stimulating nitric oxide production and vasorelaxation.

METHODS & RESULTS

Design of FRET-based biosensors for STIM1's Ca²⁺-binding loop



Figure 2. A, BS-STIM1_{3x} design (triple loop). B, Emission spectra of purified BS-STIM1_{3x} upon Ca²⁺ titration. Purified biosensors were excited at \2430 nm. C, Ca²⁺ binding curves of single-, double- and triple-loop STIM1 biosensors.

Specific interactions between EFG2 peptide and STIM1's Ca²⁺-binding loop



Figure 3. Direct interaction between EFG2 and STIM1's Ca²⁺-binding loop. A, Emission spectra of BS-STIM1_{3x} upon incremental titration of EFG2. B, Biosensor response as a function of EFG2 or Ca²⁺ concentration. C, Biosensor response to EFG2 or EFG2_{scr} N=4.



Figure 4. Effects of EFG2 on intracellular Ca²⁺ ([Ca²⁺]) in primary porcine aortic endothelial cells (PAECs) and human aortic smooth muscle cells (HASMCs). A, Effects of EFG2 on [Ca²⁺], PAECs vs HASMCs in the presence of extracellular Ca²⁺. Note the lack of effect from EFG2 on HASMCs, indicating the specificity of EFG2 to ECs.

B, Effects of EFG2 on ER Ca²⁺ stores in PAECs in the absence of extracellular Ca²⁺ followed by addition of SERCA inhibitr thapsigargin (TGN). Note the initial absence of EFG2 signal followed by the TGN-induced signal implying that the effect of EFG2 does not affect ER Ca²⁺ stores.

C, Effect of EFG2scr in PAECs in the presence of extracellular Ca²⁺. Note the absence of an effect of EFG2scr in PAECs verifying the specificity of the G2 domain in triggering Ca²⁺ entry.

Traces are averages ± SD of n = 35 – 73 cells from 3 – 6 repetitions for each condition.

G2 does not affect cardiac voltage-dependent Ca²⁺ current or cause endothelial ER stress



Figure 5. Effect of TFG2 and TFG2_{scr}on I_{CoL} in cardiomyocytes and ER stress markers IRE1 and BiP in PAECs. G2 peptide and scrambled peptide is conjugated to a universal cell penetrating sequence (TAT) to generate TFG2 and TFG2scr.

A, Current-voltage relationships of L-type Ca²⁺ current (I_{Ca}) in freshly isolated cardiomyocytes pre-injected with vehicle (CTL) or specified concentration of TFG2 and TFG2_{scr} via patch electrode tip. 5-min baseline/diffusion was allowed before data acquisition.

B, Sub-confluent PAECs were treated with the specified concentrations of TFG2 or TFG2scr before lysis and immunoblotting for the specified ER stress markers. N = 2.

In vivo vasorelaxing effects of EFG2



Figure 6. Effects of EFG2 and EFG3zer on intra-aortic blood pressure. 8-weekold male anesthetized Sprague-Dawley rats were given intravenous EFG2 or EFG2scr as arterial blood pressure was monitored. A-D, Changes in mean arterial pressure (JMAP) before and after injection of the specified peptide (arrows). N = 1 for each dose.

Loss of sustained vasorelaxing effect when G2 is tagged with a universal cell penetrating sequence



Figure 7. Effects of TFG2 on intra-aortic blood pressure. Changes in mean arterial pressure (*MMAP*) before and after injection (arrow) of TFG2 in an 8-wk-old male Sprague-Dawley rat. Note the biphasic response consisting of an initial vasorelaxation response followed by rapid vasconstriction.

SUMMARY & CONCLUSIONS

• We successfully generated a biosensor that reports interactions of binding partners with the Ca²⁺-binding loop of STIM1.

 The endothelium-specific, GPER-based EFG2 peptide directly interacts with the Ca²⁺-binding loop of STIM1 with high affinity and produces a large conformational change in this domain, whereas the control peptide with scrambled G2 sequence (EFG2scr) has a projected 100-fold lower affinity.

 In primary endothelial cells, EFG2 triggers Ca²⁺ entry while the scrambled peptide does not. EFG2 does not trigger any Ca²⁺ entry in human aortic smooth muscle cells (HASMCs), which signifies that EFG2 is selective for endothelial cells.

Using a universal cell-penetrating tag (TAT) we demonstrated that G2 does not affect voltage-dependent Ca2+ entry (in
excitable cardiomyocytes) or cause endothelial ER stress.

 In anesthetized rats, intravenous infusion of EFG2 acutely causes sustained dose-dependent vasorelaxation compared to the scrambled peptide.

When tagged to the universal cell-penetrating sequence, G2 produces a biphasic response. The initial vasorelaxation
presumably reflects NO production from the endothelium. The subsequent gradual increase in MAP likely indicates
vasoconstriction due to Ca²⁺ entry in the underlying VSMCs as TFG2 enters these cells. This response further supports the
specificity of EFG2 for the endothelium.

• These data suggest that EFG2 can be developed as a mechanistically novel therapeutic agent to improve endothelial functions in conditions such as arterial hypertension and atherosclerosis.

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